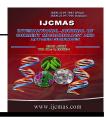
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Review Article

Post Harvest Management of Fungal Diseases in Onion - A Review

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ABSTRACT

Keywords

Onion, Post harvest losses, Fungal diseases, Management Onion is one of the highly perishable vegetable in India. It is highly valued as flouring agents. The reason for the success of onion in agriculture is its ability to be stored for long period. It can be stored up to about eight to ten months provided; it is treated accurately at its pre and post- harvest stages. About 35-40 % onion is lost due to damage caused by storage diseases. The fungal bulb rot imparts to about 15-30 % losses during storage of different varieties. There are diverse fungal pathogen species like Aspergillus spp, Penicillium spp, Alternaria spp, Fusarium spp, Rhizopus spp., Colletotrichum spp., Pseudomonas spp., Lactobacillus spp., Erwinia spp and Botrytis spp which attacks onion bulb during the post-harvest storage period. Amongst all Aspergillus spp (Especially A.niger) is the most virulent fungal pathogen in the field and during the post harvest storage. The preventive approach of using conventional fungicides to reduce the post harvest losses is not sufficient to provide safe onion and onion products for human consumption and may be hazardous for the human health due to residual impact issues. Therefore, it is essential to develop strategies to minimise the onion losses during storage by the potent application of plant derived compounds or in the form of diverse bio fungicides. The nano bio fungicides can become opportunity to counter and manage fungal diseases of onion.

Introduction

Onion is one of the potential foreign exchange earners for a country like India, as it is second largest producer of onions after China, producing 1.6 million MT annually FAO(2012). Though India ranks first in terms of the area under onion cultivation in the world and second in its production (Fig.1).The productivity is still quite low as compared to other countries Anonymous, (2001). The productivity of onion in India is 14.35 t/ha which is at least 5 times lesser as compared to republic of Korea (66.16 t/ha), about 4 times less than USA (56.13 t/ha), Spain (55.21 t/ha), Netherland (51.64 t/ha) and Myanmar (46.21 t/ha) Chengappa *et al.*, (2012). It is one of the important ingredients of the daily diet facilitating a constant year round consumer demand. It is cultivated

during kharif, late kharif and Rabi seasons as an important crop and is used in raw form as salad and also cooked as vegetable Singh *et al.*, (1994).Onion is good source of minerals, vitamins, Polyphenol and a number of phytonutrients.It lower blood pressures and prevents some kinds of cancer. There are three main seasons of onion production namely-kharif, late kharif and Rabi.

Kharif produce imparts 15-20% production and is available in the market from October to December, whereas the late kharif produce which comes in the market from January to March accounts for 20-25% production followed by the Rabi crop which is harvested in April to June which accounts for 60-65% production. The Rabi season onion is only kept for storage till October -November due to its better storability and is made available steadily for domestic as well as international markets.

The onion producing states in India includes mainly, Maharashtra, Karnataka, Gujrat, Bihar and Madhya Pradesh wherein, 32.6% of the total production is contributed only by Maharashtra, NHRDF (2012). The uncontrolled market demand with the advent of the diverse processed products of onion in the market propelled the cultivation of onion on commercial scale, which has led to a sizable increase in acreage and production .The imminent threat of storms hails and heavy rains have always been a concern for the onion growers throughout the year. Due to rains and hailstorms in Maharashtra the acreage and production of Onion declined in 2012-13 (Fig.2). Despite the achievement in production technology and availability of good varieties of onion, the post harvest losses during storage is still an ailing cause which leads to significant qualitative and quantitative losses during storage upto 25-30 %. The onion postharvest losses were estimated worth Rs 600 crores is found to be due to desiccation, decay and sprouting, Kukanoor,(2005). The rationale behind such post-harvest losses till today is the unavailability of good storage facilities during post-harvest storage phase. Their seems a big gap between the storage facility and the storage capacity which is ultimately leading to the unforeseeable postharvest decay and deterioration of onion bulbs.

The cold store capacity for fruits and vegetables in India is over 300 lakh million tonnes. Out of which most of the cold storage facilities are used for storage of onion and potato harvest and at tonnes. Post harvest losses in onion are approx Rs 1000 crores annually due to desiccation, decay, and sprouting etc, ASSOCHAM (2012). V.Anbukkarasi *et al.*, (2013) reported that during off-season the efficient storage facility for onion plays an important role for the consumers as well as for the producers which ultimately prevents serious losses due to rotting and sprouting.

Post-harvest losses in onion

Storage is one of the important aspects for post harvest handling of onion. The storage condition extends the period of availability of fresh onion by arresting the metabolic breakdown and decay. It is achieved by controlling the Relative humidity and Temperature. The storage life of onion is depends on different parameter *viz.* Physiological activity, Biochemical activity, Microbial invasion (Table1).

Inadequate and improper field curing after harvest, infection by different pathogen, sprouting and also poor storage methods being practiced by the farmers are the main reasons of prevailing losses. In general, the losses due to reduction in weight, sprouting and rotting (decay) were found to be 20-25, 4-5, and 10-12 % respectively, Panday,(1985).In India, currently about 35-40 % of the onion is estimated to be lost as postharvest losses during various postharvest operations including handling and storage, Anbukkarasi *et al.*,(2013).

Onion suffers from many diseases from pre harvest to post harvest period. They survey conducted at the international level revealed that about 35-40 % onion is lost due to damage caused by different diseases, Gupta Verma(2002). and А number of microorganisms (Table -2) are responsible for bulb rotting of onion, but among them, fungi are the main causal agent responsible for pre and post harvest period losses in the onion, Currah and proctor (1990). The tropical and sub-tropical climate regime prevailing in India, strew (1975). It makes it prone to the development of various fungal pathogens of different genera and species and in turn leading to the damage by causing rot during the storage. However, no detailed studies have been found to identify genera or species of fungal pathogens which causes rotting of onion in India.

So, Identification of pathogens which causes diseases in onion is essential for effective inhibition of target parhogens.Various of Aspergillus pathogens are species reported to cause blue mould on onion bulb during storage. The blue moulds are frequently isolated from stored diseased bulbs of local cultivars of onion. Hussain et al., (1977). Aspergillus niger is able to produce mycotoxin which reduces the quality and quantity of food products and feed-stuff which is a potent hepaticcarcinogen in humans and animals, Paster et al.,(1995); Beltmont and Carjaval, (1998); Sahin and Korukluoglu,(2000); Candlish et al.,(2001); Galvano et al.,(2001); Juglal et al.,(2002); Soliman and Badeaa, (2002); Rasooli and Abyaneh (2004); Sibi *et al.*, (2012).

The fungus causing black mold is the main member of Aspergillus and is predominantly a plant pathogen responsible for post harvest deterioration of stored food materials, Marziyeh Tolouee et al., (2010). It is responsible for the deterioration of agricultural product during pre and post harvest stages. It affects the availability of onion to domestic and international trade. The infestation of fungi causes spoilage and ultimately decreases qualitative the attributes and quantity of food (Candlish et al., 2001; Galvano et al., 2001; Soliman and Badeaa 2002. Rasooli and Abyaneh., 2004).Being saprophytic and filamentous in morphology Aspergillus niger resides and perpetuates in soil, forage, organic debris and food products causing black mold disease during post harvest stage of onion bulbs, McDonald et al., (2004).

The most favourable temperature conditions for the growth of the fungus is 28°C-34°C followed by the warm and moist conditions eliciting infection, Tysoni et al., (2004). The contamination of pathogens begins at germination stage and remains up till storage period, Hayden et al., (1994). The pathogen transmission is by infected soil or seed and the infected bulbs shows neck discoloration along with black coloured mycelia and the hiding spores in the outer dry scales Sumner et al., (1995). Chemical treatment is found best to inhibit Black mold and other fungal disease in the onion bulbs, pathogens Grinstein et al., (1992).

Preventive approaches for fungal diseases during storage

Synthetic fungicides

There are more than 50 synthetic fungicides which are recommended for reducing storage losses in onion. Various synthetic fungicides have been frequently used to prevent Pre and Post harvest fungal diseases *viz* carbendazim, Bronopol, Mancozeb, salicylic acid, Bavistin, maleic hydrazide. It was revealed the pre harvest spray of maleic hydrazide @ 2000 ppm+carbendazim@ 1000ppm at 30 days before harvest of onion bulb reduces the rotting and physiological losses and also enhances the shelf life of onion bulbs (up to six months) and improved the quality parameters like TSS content, total sugar, reducing sugar and sulphur content, Anbukkarasi, (2010).

The maleic hydrazide was banned by government of India during the year 2009 owing to its adverse affects. For the inhibition of different types of fungal pathogens there are many applications of fungicides that have been applied for a long time like Benzoyl and Carbendazim at 0.5 per cent showing better control of neck rots, Ali and Shoabrawy., (1979).

Falisolan (Carbendazim 60% + Bronopol 6%),Garcia,*et al.*,(1997) is most effective for reducing storage diseases due to *Aspergillus species*, *Fusarium species* and *Botrytis species* and also reported that disease prevalence increased drastically during third month of storage. It was also found that that post harvest spray of 0.25% mancozeb or 0.1 % carbendazim or benomyl were helpful to reduce storage losses for six month storage, Ranpise *et al.*, (2001).

Pre-harvest foliar application of Mancozeb @ 0.25 at 30 days after transplanting and repeated at fortnightly interval was adjudged better in reducing the fungal diseases and increases the onion yield in Karnal area of Haryana during Rabi season ,Anonymous, (2009).Storage losses due to fungal pathogen have been reported to be reduced by 40% by spraying of carbendazim 50% WP as pre-harvest application, Rajapakase *et al.*, (2002).However, Bose *et al.*, (2003) revealed that fumigation with sulphur dust before storage minimizes losses and sustain quality of onions in storage.

It was found that combined application of carbendazim and maleic hydrazide at 1000 and 2000 concentration ppm respectively reduced the %age of rotting, fungal infection as compared to the individual treatments in onion crop, Sable, et (2004). The harvested bulbs were al., artificially inoculated with A. niger and P. digitatum following pinprick method and stored for three months under ambient condition (27±10C). % disease reduction over control was recorded at fortnightly interval. Against black mold, carbendazim 0.1% (Bavistin) recorded maximum of 93.20 % reduction in disease 15 days after storage (DAS) and 56.91 % at 90 DAS.

Carbendazim at 0. 1% concentration was found to be most effective when applied as a post-harvest dip. In addition, post-harvest fumigation of onion bulbs with sulphur dioxide for four hours or dipping the bulbs in acetic acid at higher concentration (0.4 %)distinctly reduced the Aspergillus niger incidence, R. Srinivasan., (2006). The post harvest application of carbendazim (0.1%) was effective in controlling spoilage during storage period. The carbendazim recorded 100 % reduction in black mould and 90.7 % blue mould diseases compared to control (Raju, et al., 2007). Application of Bavistin (0.1%) proved the effective during short storage storage; whweras sulphur dust fumigation was most effective for long storage of onion, chavan et al., (1992).

The extensive use of synthetic fungicides increased the resistance of the pathogens, in addition it is not safe over public concern over food and environmental safely, therefore we have to develop the alternative methods which potentially safe to human health and environment for control the diseases (Alabi et al., 2005).

Bio fungicides

Natural bio fungicides extracted from various plant parts or microorganisms may provide an alternative to synthetic fungicides. Over the year lots of efforts has been devoted to the search for innovative antifungal materials from natural sources Karapınar,(1989);Topal,(1989); Paster et al.,(1995);De et al.,(1999); Nielsen and Rios, (2000);Galvano et al., (2001); Yin and Tsao,(2001); Juglal et al.,(2002); Soliman and Badeaa, (2002);Onyeagba et al.,(2004); Ozcan,(2005); **Boyraz** and Hacise ferogullari et al., 2005.

The application of different synthetic fungicides to control post harvest diseases of onion is a common practice but due to their toxicity and genesis of pathogen resistance facilitates the need for developing fungicidal formulations originated from plants to control fungal pathogens, Elad., (2000). Naguleswaran *et al.*, (2014) found that in a field experiment, bulb treatment together with foliar application of *Trichoderma viride* improves the yield as well as yield related parameters such as, basal diameter, circumference of bulb, mean number of bulb per bunch.

The finding is of vital importance to manage the fungal disease in red onion effectively by using plant doctor fungi *Trichoderma viride or* other *species* without the use of dangerous synthetic fungicides.

In the same year it was reported that several pathogens attack onion crop in previous section which cause many losses in yield, one of the important causal pathogens is Botrytis allii, Hussein *et al.*, (2014) .The different *Trichoderma spp.* were tested and

found to suppress the growth of *B.allii* to different degrees. The results of the antagonistic capability of *Trichoderma spp*. showed that *Trichoderma viride* caused the highest reduction of the growth compared to other species, 86%, 84% and 85% for *T*. *viride* and *T. harzianum* respectively. In case of *T. viride* as biocontrol agent it worked well under greenhouse conditions but these results are required to be confirmed by onfield study. Amongst the different fungi *Trichoderma spp*. has been reported to have greatest impact on the pathogens, McLean., (2000).

It is also reported that the crude extracts of *Moringa oleifera* has antifungal properties. They used the ethanol extracts of leaves and stems of *Moringa oleifera* to control the strain of *Aspergillus niger*. Both the plate and broth assays were used at different concentration e.g.12.5%, 25%, 50%, and 75% of extracts in Potato Dextrose Agar and Potato Dextrose Broth. The inhibitions of *Aspergillus niger* by the application of 75% concentration of leaf extract of *Moringa oleifera* Arowora *et.al.*,(2014). So, it is using as bio protective agent on onion rot which is serves as good option to chemical control.

It is reported that some pathogens also act as antagonistic effect like Penicillium species may used as biological control agents against onion fungal pathogen Aspergillus niger, Ibatsam Khokhar, et al., (2013). Penicillium roqueforti and P. viridicatum greatly inhibited the growth of A. niger by 66% and 60%, respectively. Different potential Spices antimicrobial have properties against post harvest diseases to control onion black mold using Indian culinary spices under in vitro conditions. Various plant parts of Indian cooking spices have been used for antifungal assay. It was also selected for Phyto- chemical analysis.

First round antifungal screening exposed that among fourteen spices tested, cinnamon, clove, pepper, cardamom star anise and stone flower were exhibited inhibitory activity against the black mold.

Different concentrations of the extracts ranging from 15 to 120 mg/ml were prepared and minimum inhibitory concentration (MIC) values were determined considerable inhibitory action was found at 15mg/ml concentration for cinnamon and clove. Stone flower at 30mg/ml was able to inhibit the pathogen and moderate inhibition was found in cardamom Sibi *et al.*, (2013).

From the rhizosphere of different plants species fourteen *Penicillium* species were isolated to explore the antifungal effect of *Penicillium* species against onion black rot pathogen *Aspergillus niger* Ibatsam *et al.*, (2013).After study it was found that these isolates showed very high antagonistic effect on the growth of *Aspergillus niger* mycelium .*P. viridicatum* greatly inhibited the growth of *A.niger* by 66% and 60% respectively. It was also found that *Penicillium* species completely overgrew the of *A.niger* colony.

Few Researchers investigated the phytochemical profile and effects of Plumeria against the latex postharvest fungal pathogens of sweet oranges. Post harvest fungal diseases of oranges such as Aspergillus niger, A. fumigatus, A. terreus, Penicillium digitatum and Rhizopus arrhizus were tested aligned with various extracts of Plumeria latex, Sibi et al., (2012). Antifungal effect of the extracts recorded the important inhibitory activity in opposition to Aspergillus terreus and Penicillium digitatum by the petroleum ether extract. Being for the most part effective on all species, Plumeria obtusa was create to have impending antifungal properties followed by P. rubra after five days of incubation period.

Novel preventive measures

Newer options are to be developed to find alternative options for synthetic fungicides. In a novel approach researchers reported that nanoparticles has the potential abilities to inhibit post harvest diseases of fruits and vegetables. Different types of Nanoparticles are reaching by the researchers to get the better result and take place the synthetic fungicides.

Silver nanoparticles

Othman et al., (2014) observed the effects of silver nanoparticles (AgNps) biosynthed through Aspergillus terreus (KC462061) for inhibition of growth and aflatoxin production by five isolates of A. flavus. The results showed that all five A. flavus isolates were inhibited by different concentrations of silver nanoparticles but the most excellent inhibition by 150 ppm with different significantly. Ag NPs inhibited the growth of A. flavus by disturbing cellular functions which caused deformation in fungal hyphae. AgNPs cause decrease in spores number, abnormality and hypertrophy, these special effects lead to destroyed and damaged of spores. Gajbhiye et al. (2009) revealed that combination of nano particles of silver and antifungal agents like fluconazole can increase antifungal effectiveness of disinfectants.

The function of microorganisms in the synthesis of silver nanoparticles (AgNps) emerges as an eco-friendly and sustainable approach. San et al., (2013) reported the antimicrobial property of silver nanoparticles synthesised through reduction with mycelia and culture of Schizo phylum and silver nitrate. The antimicrobial activities of silver nanoparticles are tested against Aspergillus niger, Staphylococcus *Staphylococcus* epidermidis, aureus.

Escherichia coli, and Candida albicans. Results showed that silver nanoparticles synthesised by interaction silver nitrate with mycelia fungus, when treated with **Staphylococcus** aureus and silver nanoparticles synthesised by interaction of silver nitrate and culture supernatant treated with Staphylococcus epidermidis, gave the largest inhibition area.

In the same year, Khadri et al., (2013) develop potential anti-microbial particles of silver nanoparticles by green synthesis an approach conventional eco-friendly to synthesis. chemical The enzymatic mechanism of the olive seeds has been subjected to generate silver nanoparticles and test the efficacy as antifungal agents before characterizing physical properties using FTIR, UV-Vis and TEM analysis. Silver nano particles were proven to inhibit the growth and development of mycelia of Aspergillus niger.

The antifungal activity of composite films, nanoparticles Ag (NP) and pullulan against Aspergillus These niger. new materials were prepared as transparent cast films (66–74 µm thickness) from Ag hydrosols containing the polysaccharide Pinto et al., (2012). Fungal growth inhibition was observed in the incidence of such silver nanocomposite films. Moreover. interruption of the spores' cells of A. *niger* was probed for the first time by means of scanning electron microscopy (SEM).

Lamsal et al., (2011) reported the effect of nanoparticles silver against pepper anthracnose under different concentrations and silver nanoparticles were applied at different concentrations to determine the in vitro antifungal effects and simultaneously in the field conditions. The application of 100 ppm concentration of silver nanoparticles created maximum inhibition of conidial germination as well as the growth of fungal hyphae in comparison to control under *in vitro* conditions and under the field conditions showed inhibition of fungal attack before disease outbreak on the plants. The application of silver nanoparticles in the liquid formulation is found to have inhibiting affect at a concentration of 7 ppm against the White Rot of green onion *Sclerotium cepivorum* (Jin-Hee Jung.,(2010).

Copper nanoparticles

Copper nanoparticles have been reported an important role in pathogen inhibition, Prachi *et al.*, (2014). It is reported that antifungal activity of copper nanoparticles against *Fusarium oxysporium* and found antifungal activity of bavistin increases in combination with CuNPs in the cases of *Fusarium oxysporium*. In the same year the antifungal activity of silver (AgNPs), copper (CuNPs) and silver/copper (Ag/CuNPs) nanoparticles against two plant pathogenic fungi *Botrytis cinera* and *Alternaria alternata*, Sahar (*et al.*,2014).

Metal nanoparticles were applied at different concentrations to determine antifungal activities in vitro. The application of 15 mg L⁻¹ concentration of silver nanoparticles gave maximum inhibition of the growth of fungal hyphae. assessed They the effectiveness of combining the silver and copper nanoparticles. Microscopic study revealed that nanoparticles caused a damage effect on conidia and fungal hyphae. It was reported the antifungal efficiency of Zinc oxide nanoparticles (ZnO NPs) against two pathogenic fungal species, Fusarium oxysporum and Penicillium expansum. The antifungal activity of ZnO NPs was found to be concentration dependent, Ramy et al., (2013) .Hence, maximal inhibition of Mycelial growth corresponded to the highest

experimental concentration (12 mg L^{-1}), where 77 and 100% growth inhibition was observed for *F. oxysporum* and *P. expansum*, respectively.

Sulphur and Zinc oxide nanoparticles

Massalimov et al., (2013) investigated the antifungal effect of micron and nano scale form of sulphur against two types of pathogenic Schutte fungi. The antifungal effects of sulphur particles in field experiment and sabouraud medium have been studied. It was found that in all cases of laboratory as well as field experiment, antifungal activity of nano sized sulphur with an average particle size of 25 nm, 5-10 times higher than the sulphur micron with an average particle size of 8 microns. Choudhury et al., (2010) synthesizes Sulphur nanoparticles (NPs) via a liquid synthesis method with the particles size in the range of 50-80 nm in spherical shape. A comparative study of elemental and nano sulphur was done against the inhibition of Aspergillus niger. The results showed that nano sulphur is more effective against Aspergillus niger than its elemental form.

Srinivasan *et al.*, (2015) investigated the antifungal performance of Zinc oxide nanoparticles against *Aspergillus nigar and Aspergillus flavus*. The pure zinc oxide nanoparticles were found to be efficient fungicide against *Aspergillus niger* but the aluminium droped zinc oxide nanoparticles were active against *Aspergillus flavus*. The zone of Inhibition diameter value of aluminium doped zinc oxide against *Aspergillus flavus* was found maximum 13 mm for 100μ g/ml whereas the zone of Inhibition diameter value of against *Aspergillus nigar* was found maximum 11 mm for 100μ g/ml.

Navale *et al.*, (2015) investigated the antimicrobial activities of Zinc oxide

nanoparticles (ZnO NPs) against two strains of plant pathogenic fungi Aspergillus flavus and Aspergillus fumigatus. These antimicrobial data indicates that the ZnO NPs (Size 20-25 nm) are potentially active in the presence of 100 µg/mL NPs. Singh et al., (2013) Claimed that zinc oxide nanoparticles do have strong good antifungal activity against selected strains of bacteria and fungus as compared to that of conventional zinc oxide particles. ZnO NPs have been claimed to have pronounced antimicrobial activities than large particles. Antimicrobial/antifungal potential of ZnO on five pathogens (Escherichia coli, Staphylococcus aureus, Bacillus subtilis, Aspergillus niger and Candida albicans) and the influence of particles size of these inorganic powders on its antimicrobial /antifungal efficacy were considered in the present study.

Yehia et al., (2013) investigated the antifungal effectiveness of Zinc oxide nanoparticles (ZnO NPs) against two pathogenic fungal species, Fusarium oxysporum and P. expansum. The maximal zone of inhibition of mycelial growth corresponded to the highest experimental concentration (12mgL^{-1}) , where 77 and 100% growth inhibition was observed for F. oxysporum and P. expansum, respectively. The antifungal activity of ZnO NPs was found concentration dependent.Wani et al.,(2012) studied the antifungal activity of nanoparticles of magnesium, Iron and zinc. It had been investigated under in-vitro conditions and was found that all the nanoparticles at different concentrations brought about significant inhibition in the germination of spores of Aspergillus niger, Penicillium notatum, and Nigrospora oryzae Berk. However, the highest inhibition in the germination of all the test fungi was observed at higher concentrations followed by lower concentrations of nanoparticles.

Table.1 Different factors impacting the post harvest losses in onion

Physiological activity	Biochemical activity	Microbial invasion
1. Transpiration	1.Enzymatic	1.Fungi
2. Respiration	2.Softeninf of tissues	2. Bacteria
3. Senescence		
4. sprouting		

Table.2 Source: Onion disease Guide- Seminis® is a registered trademark of Seminis Vegetable

 Seeds. Inc. ©2012 Seminis Vegetable Seeds, Inc.

Sl No.	Name of disease	Causal agents	Distribution
1.	Basal Rot	Fusarium oxysporum f. sp. cepae	Worldwide
2.	Black Mold	Aspergillus niger	Worldwide
3.	Black Stalk Rot	Stemphylium botryosum	Worldwide
4.	Blue Mold Rot	Penicillium species	Worldwide
5.	Botrytis Brown Stain	Botrytis cinerea	North America and Europe
6.	Botrytis Leaf Blight	Botrytis squamosa	North America and Europe
7.	Damping-Off	Fusarium species, Pythium species, Rhizoctonia solani	Worldwide
8.	Downy Mildew	Peronospora destructor	Disease occurs worldwide in temperate and cool growing regions
9.	Leaf Blotch	Cladosporium allii-cepa	British Isles and Canada
10.	Neck Rot	Botrytis allii	Worldwide
11.	Phytophthora Neck And Bulb Rot	Phytophthora nicotianae	Brazil and Taiwan
12.	Pink Root	Phoma terrestris	Worldwide
13.	Powdery Mildew	Leveillula taurica	Brazil, Israel, Italy, Turkey and USA
14.	Purple Blotch	Alternaria porri	Worldwide
15.	Rust	Puccinia allii	Disease occurs worldwide in temperate and cool growing regions.
16.	Smudge	Colletotrichum circinans	Worldwide
17.	Smut	Urocystis colchici,	Worldwide
18.	Southern Blight	Sclerotium rolfsii	Worldwide
19.	Stemphylium Leaf Blight	Stemphylium vesicarium	India and USA, however, the pathogen may occur in other onion growing regions of the world.
20.	Twister	Colletotrichum gloeosporioides	Worldwide, although only of significance in tropical and sub-tropical regions.
21.	White Rot	Sclerotium cepivorum	Worldwide
22.	White Tip	Phytophthora porri	Worldwide
23.	Yeast Soft Rot	Kluyveromyces marxianus var. Marxianus	USA

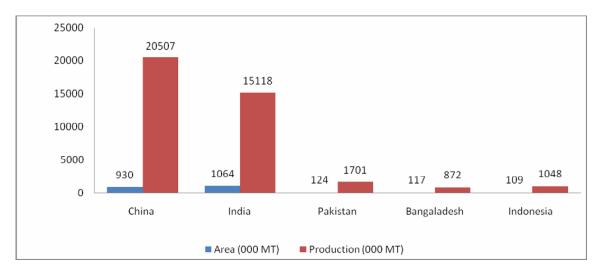
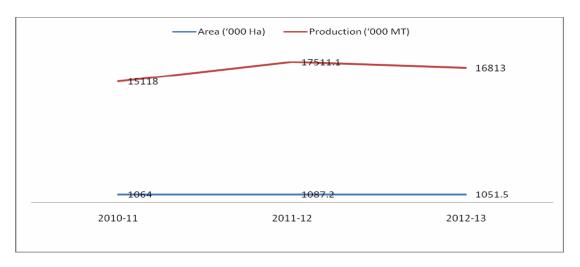


Fig.1 Production of Onion and area under cultivation in world (Source: Indian Horticulture Data base, 2013)

Fig.2 Production of Onion and area under cultivation in India (Source: Horticulture Statistics Division, D/o Agriculture & Cooperation, 2013)



The Nano-MgO at highest concentration was found most effective in reducing the spore germination followed by Nano-FeO and Nano-ZnO at the same concentration respectively. Zinc oxide nanoparticles (ZnO NPs) at the concentration greater than 3moll⁻¹ are reported to have significant antifungal activity against the two post harvest fungal pathogens i.e. *Botrytis cinera and Penicillium expansum* during storage, Lili *et*

al., (2011).The growth of conidiophores and conidia of *P. expansum* are prevented by ZnONPs ultimately leading to the death of fungal pathogen. Nanoparticles have the important role in controlling the diverse post harvest disease pathogenicity. Researchers are trying to find out diverse nanoparticles for specific post harvest diseases of onion and vegetables crop.

Knowledge gaps

Now days some of the knowledge gaps are present which were identified during this review and there is urgent need for further research are as follows:

- 1. Limited knowledge is available on application of bio fungicide in Indian farmers.
- 2. Only *in vitro* results are available and it is not used by the farmers on large scale.
- 3. Lack of long-term post harvest studies limits the understanding bio fungicides and plant based bio- products for commercial use.
- 4. No standard application rate of bio fungicide for post harvest diseases management on onion to get maximum significant positive results is available.
- 5. Farmers have faith only on the use of synthetic fungicide for controlling post harvest diseases of onion.

There is tremendous need for developing environmentally friendly fungicidal formulations against the fungal pathogens worldwide to cope up with the toxic residual impacts of traditional synthetic fungicides on the human health. Diverse techniques for the identification of fungal pathogens must be developed so, as to target the pathogen ailing the Onion bulbs at post-harvest phase. Various fungicidal formulations should be especially bio-fungicidal used the formulations to prevent any type of health hazard to the consumers. The prevailing present challenge is to develop strategies to identify location and region based ecotypes/pathotype sustaining at different geographical locations and accordingly the ways to tackle the post-harvest rotting and decay could be managed by formulating the bio fungicides targeting the region prevalent pathogens successfully. Nanotechnology could also impart in developing ecofriendly, sustainable methodology to mitigate post-harvest losses in onion.

References

- A. H. Wani, M. Amin, M. Shahnaz and M.
 A. Shah 2012, Antimycotic Activity of Nanoparticles of MgO, FeO and ZnO on some Pathogenic Fungi. International Journal of Manufacturing, Materials, and Mechanical Engineering (IJMMME) -2(4):12
- Alabi, D., I Oyero and N. Amusa, 2005, Fungitoxic and phytotoxic effect of amygdalina Vernonia (L.), pinnantus Bryophyllum Kurz Ocimum gratissimum (Closium) L. and Eucalyptna globules (Caliptos) Labill water extracts on cowpea and cowpea seedling pathogens in Ago-Iwoye, South Western Nigeria. World Journal of Agricultural Sciences, pp: 1.
- Ali, A.A., et al, 1979, Effect of some cultural practices and chemicals on the control of neck rots diseases caused by *Botrytis allii* during storage. Agricultural Research Review, 52(2): 103-104.
- Anbukkasari, V. 2010, Studies on pre and post harvest treatments for extending shelf life of onion (*Allium cepa* L.var aggregatum don).cv Co on 5. Ph.D. Thesis, department of vegetable crops, Tamilnadu agriculture university, Coimbatore.
- Anonymous, 2001. Annual progress report for kharif, 1998, Division of Plant Pathology, S. K. University of Agriculture Science and Technology (Kashmir), 34 pp.

- Anonymous, 2009, National Horticultural Research and Development Foundation (NHRDF), Nasik-Newsletter, XXIX (4).
- Arowora K.A. and Adetunji C.O 2014, Antifungal effects of crude extracts of Moring oleifera on *Aspergillus niger* v. Tieghem associated with post harvest rot of onion bulb. SMU medical journal 1(2) 2014-223.
- Belmont RM, Carjaval M, 1998, Control of *Aspergillus flavus* in maize with plant essential oils and their components. J. Food Prot. 61:616-619.
- Bose, T.K., *et al.*, 2003, *Vegetable Crops*. Naya Udyog Publisher, Kolkata, 3: 68-69.
- Boyraz N, Özcan M 2005, Antifungal effect of some spice hydrosols. Fitoterapia. 76: 661- 665.
- Candlish AAG,Pearson SM,Aidoo KE,Smith JE, Kell B and Irvine, H. 2001, A survey of ethnic foods for microbial quality and aflatoxin content. Food Addit Contaminants. 18: 129-136.
- Chan Yen San and Mashitah Mat Don., 2013, Biosynthesis of Silver nanoparticles from *Schizophyllum* Commune and In-vitro Antibacterial and Antifungal Activity Studies. Journal of Physical Science, 24 (2): 83–96
- Chavan, V.B., T.F. D'Souza, S.B. Kokate and D.M. Sawant.,1992, Efficacy of chemicals in restricting onion bulb rots in storage. Maharashtra Journal of Horticulture. 6(2): 92-94.
- Chengappa, P.G., Manjunatha, A.V., Vikas Dimble, Khalil shah., 2012, Competitive Assessment of Onion Markets in India. Institute For Social and Economic Change.Competition Commission of India.1-86.

- Currah, L. and F.J. Proctor.,1990, Onion in tropical region. Bulletin no.35, Natural Resources Institute, Chatham, Maritime, Kent, UK, 79p.
- Elad, Y., 2000, Biological control of foliar pathogens by means of *Trichoderma harzianum* and potential modes of action. Crop Protection, 19: 709.
- FAO. Onion Production. Food and Agriculture Organization (FAO) of the United Nations, FAOSTAT. 2012, <u>http://faostat.fao.org</u>.
- Gajbhiye M., Kesharwani J., Ingle A., Gade A., Rai M., 2009, Fungus-mediated synthesis of silver nanoparticles and their activity against pathogenic fungi in combination with fluconazole. Nanomedicine 5, 382– 386.
- Galvano F,Piva A, Ritieni A,Galvano G., 2001,Dietary strategies to counteract the effects of mycotoxin: Rev. J. Food Protect. 64:10-131.
- Galvano, F., Piva, A., Ritieni, A., and Galvano, G.,2001, Dietary strategies to counteract the effects of mycotoxin: Rev. Journal of Food Protection 64:10-31.
- Garcia, J.L., Lopez, M.T., Perera, E.T., Berkmunn, H. and Gonzalez, L.A., 1997, Investigations on chemical control during storage of onion bulbs. Agrotecnia-de -Cuba, 27(1): 8-10.
- Govinda R Navale, Thripuranthaka M, Dattatray J Late and Sandip S Shinde.,2015, Antimicrobial Activity of ZnO Nanoparticles against Pathogenic Bacteria and Fungi. JSM Nanotechnology & Nanomedicine. 3(1): 1033.
- Grinstein A, Elad Y, Temkin GN, Rivan Y, Frankel H., 1992, Reduced volume application of fungicides for the control of onion rots. Phytoparasitica; 20: 293-300.

- Gupta, R.P.and Verma, L.R., 2002, Problem of diseases during storage in onion and garlic and their strategic management. In implication of plant diseases on produce quality (Eds.singh, D.P.).Pp.55-62.kalyani publishers, Ludhiana.
- Habeeb khadri, Mohammad alzohairy, Avilala janardhan, Arthala Praveen Kumar and Golla Narasimha., 2013, Green Synthesis of Silver Nanoparticles with High Fungicidal Activity from Olive Seed Extract. Advances in Nanoparticles. 2: 241-246.
- Haciseferogulları H, Özcan M, Demir F, Calısır S.,2005, Some nutritional and technological properties of garlic (*Allium sativum* L.). J. Food Eng. 68: 463-469.
- Hayden NJ, Maude RB, Proctor FJ., 1994, Studies on the biology of black mould (*Aspergillus niger*) on temperate and tropical onions. 1. A comparison of sources of the disease in temperate and tropical field crops. Plant Path; 43: 562-569.
- Hussain, F.N., Abd-Elrazik, F.A., Darweish arj, and Rushdi, M.H., 1977, Survey of storage diseases of onion and their incidents in upper Egypt.J.phytopath9:15.
- Ibatsam Khokhar, Muhammad Saleem Haider, Irum Mukhtar, Sobia Mushtaq., 2013, Biological control of *Aspergillus niger* the cause of Black-rot disease of (*Allium cepa* L).Onion by Penicillium species. Journal of Agrobiology 29(1): 23– 28.
- Jin-Hee Jung, Sang-Woo Kim, Ji-Seon Min, Young-Jae Kim, Kabir Lamsal and Kyoung Su Kim.,2010, The Effect of Nano-Silver Liquid against the White Rot of the Green Onion

Caused by Sclerotium cepivorum. Mycobiology.38 (1): 39–45.

- Juglal S, Govinden R, Odhav B., 2002, Spice oils for the control of cooccurring mycotoxin producing fungi.J. Food Protection. 65: 683-687.
- Kabir Lamsal, Sang Woo Kim, Jin Hee Jung, Yun Seok Kim, Kyong Su Kim and Youn Su Lee.,2011, Application of Silver nanoparticles for the control of *Colletotrichum Species* In vitro and Pepper Anthracnose Disease in Field. Microbiology 39(3): 194-199.
- Karapınar M., 1989, Inhibition effects of some spice agents on aflatoxi- genic mould growths. International Food Symposium.4-6 April, Bursa Turkey, Proc. Book.129- 137.
- Kukanoor, 2005, Post–harvest studies in onion Cv.N-53. PhD. University of Agriculture sciences, Dhawan.
- Lili He, Yang Liu, Azlin Mustapha and Mengshi Lin., 2011, Antifungal activity of zinc oxide nanoparticles against *Botrytis cinerea* and *Penicillium expansum*. Microbiological Research.166 (3):207–215.
- M. R. Al Othman, A. R. M. Abd El Aziza,
 M. A. Mahmoud, S. A. Eifan, M. S. El Shikh, M. Majrashi.,2014,
 Application of Silver nanoparticles as Antifungal and antiaflatoxin B1
 Produced by Aspergillus Flavus.
 Digest Journal of Nanomaterials and
 Bio structures. 9(1)151 157.
- M.A.M. Hussein, M.H.A. Hassan and K.A.M. Abo-Elyousr., 2014, Biological Control of Botrytis allii by Trichoderma viride on Onion Allium cepa. World Applied Sciences Journal 32 (3): 522-526.
- Marziyeh Tolouee, Soheil Alinezhad, Reza Saberi, Ali Eslamifar, Seyed Javad

Zad, Kamkar Jaimand, Jaleh Taeb, Mohammad **Bagher** Rezaee. Masanobu Kawachi, Masoomeh Shams Ghahfarokhi, Mehdi Razzaghi Abyaneh.,2010, Effect of Matricaria chamomilla L.flower essential oil on the growth and ultra structure of Aspergillus niger van Tieghem. International Journal of Food Microbiology, 139: 127–133.

- Massalimov I.A., Zaynitdinova R.M., Shaynurova A.R. and Mustafin A.G.,2013, The efficacy of Micron and nanoscale Sulphur the Schutte Fungi. International Journal of Sciences.2:27-30.
- McDonald MR. Jaime MA. Hovius Management MHY.,2004, of diseases of onions and garlic. In Diseases of fruits and vegetables. Eds. Naqvi SAMH. Kluwer Academic The Publishers. Netherlandspp: 149-200.
- McLean, K and A. Stewart, 2000, Application strategies for control of onion white rot by fungal antagonists. New Zealand Journal of Crop and Horticultural Science, 28: 115
- N. Srinivasan, J. C. Kannan and S. Sathees kumar., 2015), Antifungal activity of pure and aluminium doped zinc oxide nanoparticles against *Aspergillus nigar* and *Aspergillus flavus*. International Journal of Pharm Tech Research. 7(2): 287-290.
- Nielsen PV, Rios R.,2000, Inhibition of fungal growth on bread by volatile components from spices and herbs ,and the possible application in active packaging, with special emphasis on mustard essential oil. Int. J. Food Microbiol. 60:219-229.
- Onyeagba RA, Ugbogu OC, Okeke CU, Iroakasi O., 2004, Studies on the

antimicrobial effects of garlic (*Allium sativum* Linn), ginger (*Zingiber officinale* Roscoe) and lime (*Citrus aurantifolia* Linn). Afr. J.Biotechnol. 3:552-554.

- Pandey, U.B., 1989, Problems in postharvest handling of onion and current status of research work done by AADF in the field of post-harvest technology. AADF News Letter, 9(3 & 4): 12-15.
- Paster N, Menasherov M, Ravid U, Juven B.,1995, Antifungal activity of oregano and thyme essential oils applied as fumigants against fungi attacking stored grain. J. Food protects. 58:81-85.
- Priyanka Singh and Arun Nanda.,2013, Antimicrobial and antifungal potential of zinc oxide nanoparticles in comparison to conventional zinc oxide particles. Journal of Chemical and Pharmaceutical Research. 5(11):457-463.
- Rajapakse, R.G.A.S. and Edirimanna and E.R.S.P., 2002, Management of bulb rot of big onion (*Allium cepa* L.) during storage using fungicides. Annals of the Sri Lanka department of agriculture, 4:319-326.
- Raju, K., and M.K. Naik., 2007, Effect of post-harvest treatments of onion to control spoilage during storage. Journal of Food Science Technology, 44(6): 595-599.
- Ramy S. Yehia and Osama F. Ahmed., 2013, In vitro study of the antifungal efficacy of zinc oxide nanoparticles against *Fusarium oxysporum* and *Penicilium expansum*. African Journal of Microbiology Research. 7(19) 1917-1923.
- Ranpise, S.A., R.M. Birade, B.T. Patil and S.V. Swant., 2001, Factors affecting the storage of onion: A review.

Orissa Journal of Horticulture, 29(1): 1-12.

- Rasooli I, Abyaneh MR., 2004, Inhibitory effects of thyme oils on growth and aflatoxin production by Aspergillus parasiticus. Food Control. 15: 479-483.
- Renu Gupta, MK Khokhar and Ram Lal., 2012, Management of the Black Mould Disease of Onion. Journal of Plant Patho Microbes, 5(3):312-315.
- Ricardo J.B. Pinto, Adelaide Almeida, Susana C.M. Fernandes, Carmen S.R. Freire, Armando J.D. Silvestre, Carlos Pascoal Neto, Tito Trindade.,2012, Antifungal activity of transparent nanocomposite thin films of pollutant and silver against *Aspergillus niger*. Colloids and Surfaces B: Biointerfaces. (103): 143-148.
- Sabale, A. and S. Kalebere.,2004, Storage behaviour of onion (*Allium cepa* L.) varieties under the influence of preharvest and post-harvest treatment of maleic hydrazide and carbendazim. Acta Botanica Hungarica., 46(34):395–400.
- Sahar M Ouda, 2014, Antifungal activity of silver and copper nanoparticles on two plant pathogens, Alternaria alternata and Botrytis cinera. Research journal of microbiology.9 (1):34:43.
- Sahin I, Korukluoglu M., 2000, Mould-Food-Human.Uludag University Pres,Vipas, Bursa. Pp. 3-122 (In Turkish).
- Samrat Roy Choudhury, Kishore K. Nair, Rajesh Kumar, Robin Gogoi, Chitra Srivastava, Madhuban Gopal, B. S. Subhramanyam, C.devakumar and Arunav Goswami .,2010,International conference on advanced Nanomaterials and

nanotechnology. American Institute of Physics.154-157.

- Sibi G, Apsara V, K. Dhananjaya,H. Mallesha and K. R. Ravi kumar .,2012, Biological control of postharvest fungal pathogens of sweet oranges by Plumeria latex. Asian Journal of Plant Science and Research 2 (5):613-619
- Sibi G, Awasthi S, Dhananjaya K, Mallesha H, Ravi kumar KR., 2012, Comparative studies of Plumeria species for their phyto-chemical and antifungal properties against Citrus sinensis pathogens. Int J Agri Res; 7(6): 324-331.
- Sibi G, Rashmi Wadhavan, Sneha Singh, K. Dhananjaya, K.R. Ravi kumar And H. Mallesha.,2013, Biological control of onion black mold by Indian culinary spices under *in vitro* conditions. Asian Journal of Pharmaceutical and Clinical Research .6(2): 156-158
- Singh S, Singh AP, Sinha S.B., 1994. Effect of spacing and various levels of nitrogen on seed crops of kharif onion, Veg. Sci., 21: 1-6.
- Soliman KM, Badeaa RI., 2002, Effect of oil extracted from some medicinal plants on different myco-toxigenic fungi. Food Chem. Toxicol. 40: 1669-1675.
- Srinivasan, R. and Shanmugam, V., 2006, Post harvest management of black mould rot of onion. Indian Phytopathology.59 (3): 333-339.
- Strew, J.R., 1975, Effect of humidity on losses of bulb onion (Allium cepa L) stored at high temperature.Expt.Agril11:81.
- Sumner D.R., 1995, Diseases of bulbs caused by fungi -Black mold. In: F. Schwartz and S.K. Mohan., (Eds). Compendium of onion and garlic

disease. APS press. St. Paul, Minn.; pp. 26-27

- Topal S., 1989, The researches about antimicrobial effects of garlic and onion. International Food Symposium.4-6 April, Bursa Turkey, Proceeding Book. pp 450-462 (In Turkish).
- Tsao SH, Yin MC., 2001, In vitro antimicrobial activity of four diallylsulphides occurring naturally in garlic and Chinese leek oils. J. Med. Microbiol. 50:646-649.
- Tysoni JL, Fullerton RA., 2004, Effect of soil borne inoculums on incidence of onion black mold (*Aspergillus niger*). New Zealand Plant Protection; 57: 138-141.
- V.Anbukkarasi,P.Paramaguru L.Pugalendhi, N. Ragupathi and P. Jeyakumar, 2013, Studies on pre and postharvest treatments for extending shelf life in onion – A review, Agricultural research communication centre., 34 (4) : 256-268.
- Vigitha Naguleswaran, Kandiah Pakeerathan and Gunasingam Mikunthan., 2014, Biological Control: A Promising Tool for Bulb-Rot and Leaf Twisting Fungal Diseases in Red Onion (*Allium Cepa* L.) In Jaffna District. World Applied Sciences Journal 31 (6): 1090-109.